

Dutta, S. K. Studier on nucleic acid interactions and chromatin isolated from differentiated cells.

stitution, Washington, D.C. and D. P. Bloch of the Institute of Cell Research, University of Texas, Austin, Texas. **These** techniques have made the following studier possible:

(1) Studier on repeated DNA sequence in N. crassa. While most **eucaryotic** organisms contain large numbers of repeated DNA sequences, N. crassa has very few (Dutta and Kohne 1969 **Proc. XI Intern. Botany Congr. 1969:50**), if any, of such **repeated sequences**. **Approximately** 10% of the whole cell DNA is found to be repeated. This is believed to be **mostly mitochondrial** DNA. This will be an extremely useful property in the interpretation of the nucleic acid hybridization data. Furthermore, it has been possible to study the entire kinetics of DNA reassociation. **This** knowledge enabler an accurate measurement, within 1% error, of the identity of **nucleotide** sequencer of DNA from different cell types. Comparisons of half Cot values (Cot = $(OD \text{ at } 260 \text{ m}\mu/2) \times \text{hours of incubation}$: $1/2 \text{ Cot} = \text{Cot value for } 50\% \text{ hybridization}$: **Britten and Kohne 1968 Science 161:529**) of E. coli (standard) DNA with N. crassa DNA enable us to conclude that the "information content" of N. crassa nuclear DNA is close to 2×10^{10} daltons. This indicates that N. crassa nuclear DNA will take 15 hours, in comparison with 750 hours for DNA of the cow, in order to get 95% **DNA:DNA reassociation** at a concentration of 5 mg DNA/ml in 0.18 M sodium ion. Bored on the some technique, we have found that the information content of **Neurospora mitochondrial** DNA is 7×10^7 and that there are only 30 copier of DNA repeats per cell.

(2) Studies on differential gene expression by DNA:RNA hybridizations. The earlier studier made with higher organisms on this problem are based on **DNA-agar** and membrane filter techniques measuring only the expression of repeated sequencer of DNA Using these techniques, we hove not been able to **obtain** more than 30% DNA:DNA hybridization compared with the 98% easily obtained by the **hydroxyapatite** technique (**Britten and Kohne ibid.**) between the identical DNAs. It should be possible to **isolate** RNA **cistrans** from different cell types of **Neurospora** by this technique, using the procedure of Kohne (1968 **Biophys. J. 8: 1104**).

(4) Studies on chromatinr isolated from differentiated cells of N. crassa. Several workers have established the usefulness of the study of the chemistry of **chromatins** for understanding the molecular **basis** of **morphogenesis** in higher organisms. **Our** studier regarding the chemical composition of chromatinr and basic proteins (**Dwivedi, Dutta and Bloch 1969 J. Cell Biol. 43:51**) in-

Recently we have developed techniques of DNA:DNA and DNA:RNA hybridization and of **chromatin** isolation permitting studies on a molecular **basis** of differentiation in **Neurospora**, in collaboration with **D. E. Kohne** of the Department of Terrestrial Magnetism, Carnegie In-

dicate that probably some different kind of **basic** proteins (other **than** any known **histones**) are involved in such **lower eucaryotic** organisms. We have **shown** (**Dutta** and Crockett 1968 **The Nucleus**, p. **65**, Calcutta Univ. **Seminar Vol.**) that there are **some** differences in chemical constituents of DNA and RNA in **chromatins** isolated from **mycelial** and **conidial** cells.

All of **these** studies indicate very strongly **the** value of working with **Neurospora** cell types and morphological **mutants** to gain useful knowledge **regarding** the molecular basis of differentiation. Part of these **studies** are already **published**, and parts **are** in the **process** of **publication** elsewhere. This research **has** been supported by a NSF grant **GY 3894**. • • • Department of Botany, Howard **University**, Washington, D.C. **20001**.