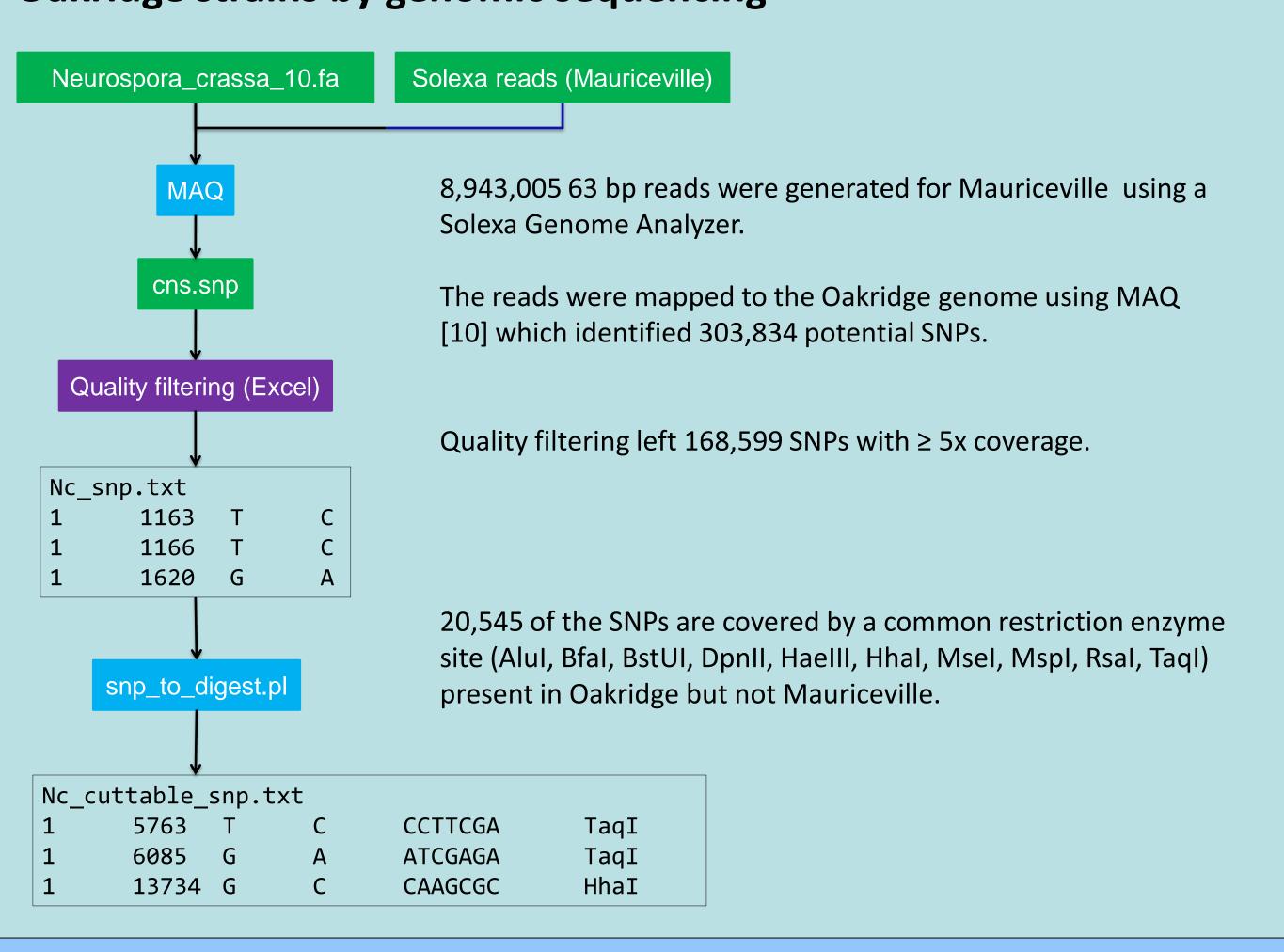


A SNP map for Neurospora crassa Mauriceville

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Identification of SNPs between the N. crassa Mauriceville and Oakridge strains by genomic sequencing



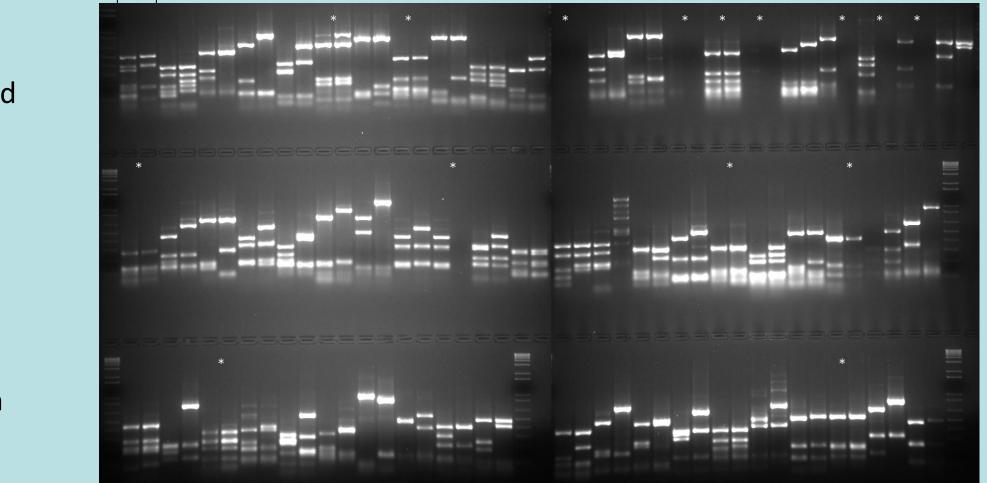
SNP validation

1) SNPs in Mauriceville EST data

A verified C to G SNP 95/100 SNPs screened are Mauriceville: 193 agagatgtgtattggtgtcaaagggcaatgactgatgattc 115 present in a BLAT aligned Mauriceville EST dataset. Oakridge: 119025 agagatctgtattggtgtcaaagggcaatgactgatgattc 119065

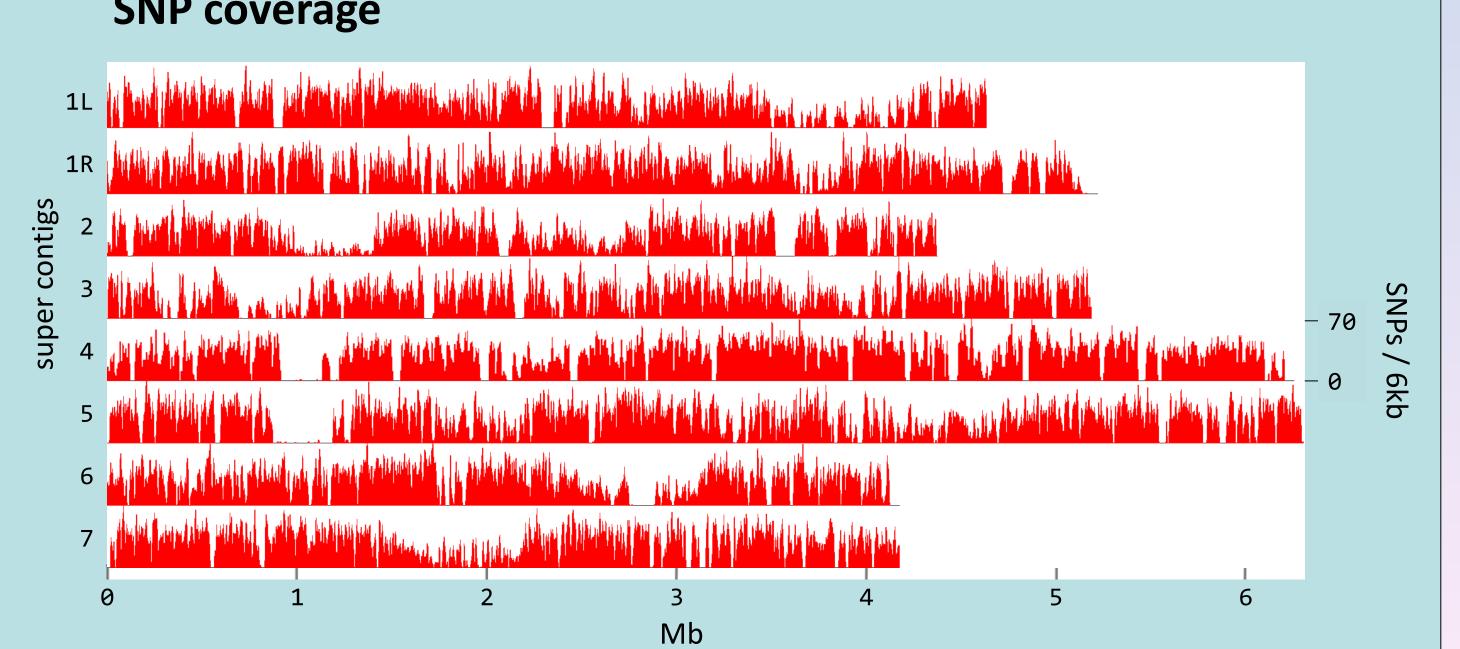
2) RFLP patterns after digesting PCR products covering Taql digestible SNPs

48/48 SNP containing regions amplified by PCR from Mauriceville (M) and Oakridge (O) have the expected RFLP pattern.



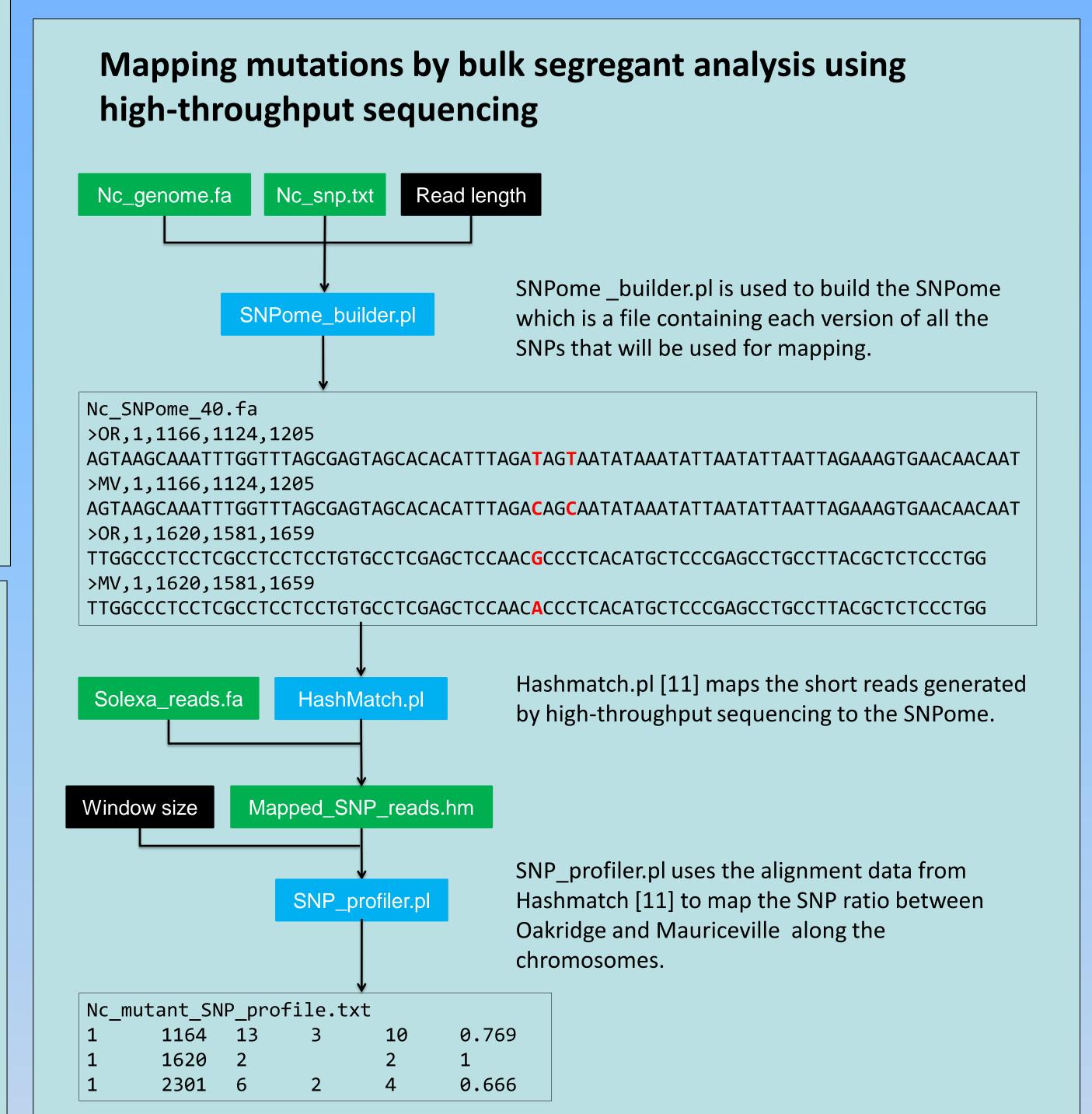
*Ambiguous RFLP where the greatest difference in band size is < 10bp.

SNP coverage

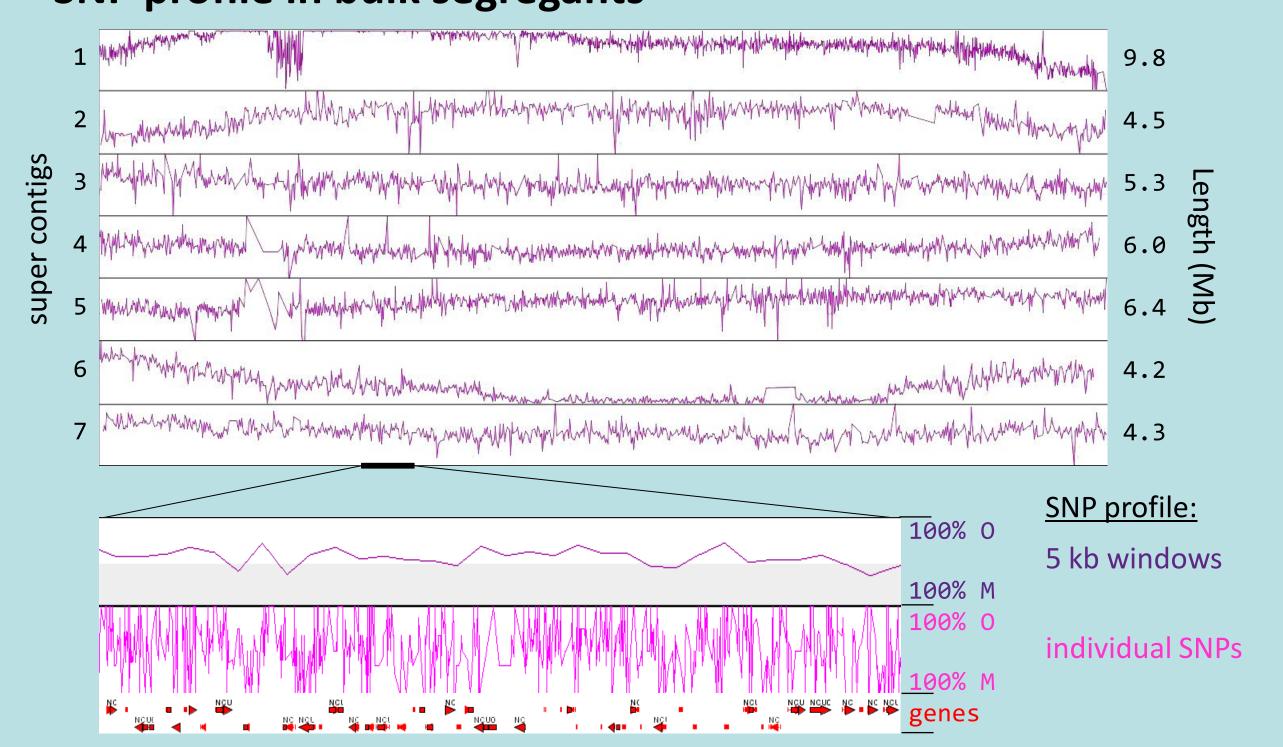


Abstract

With the advent of high-throughput DNA sequencing it is now straight-forward and cheap to generate high-density small nucleotide polymorphism (SNP) maps that can improve on mapping strategies based on RFLP [1], CAPS [2,3] or RAD-mapping [4,5] analyses. Direct identification of single point mutations has been described in fission yeast [6] but in most organisms bulk segregant analyses followed by SNP mapping are used [7]. Here we present a high density SNP map between Neurospora crassa Mauriceville-1-c (FGSC2225) [8] and OR74A (FGSC987) [9], the strains most typically used by Neurospora researchers to carry out RFLP mapping crosses.

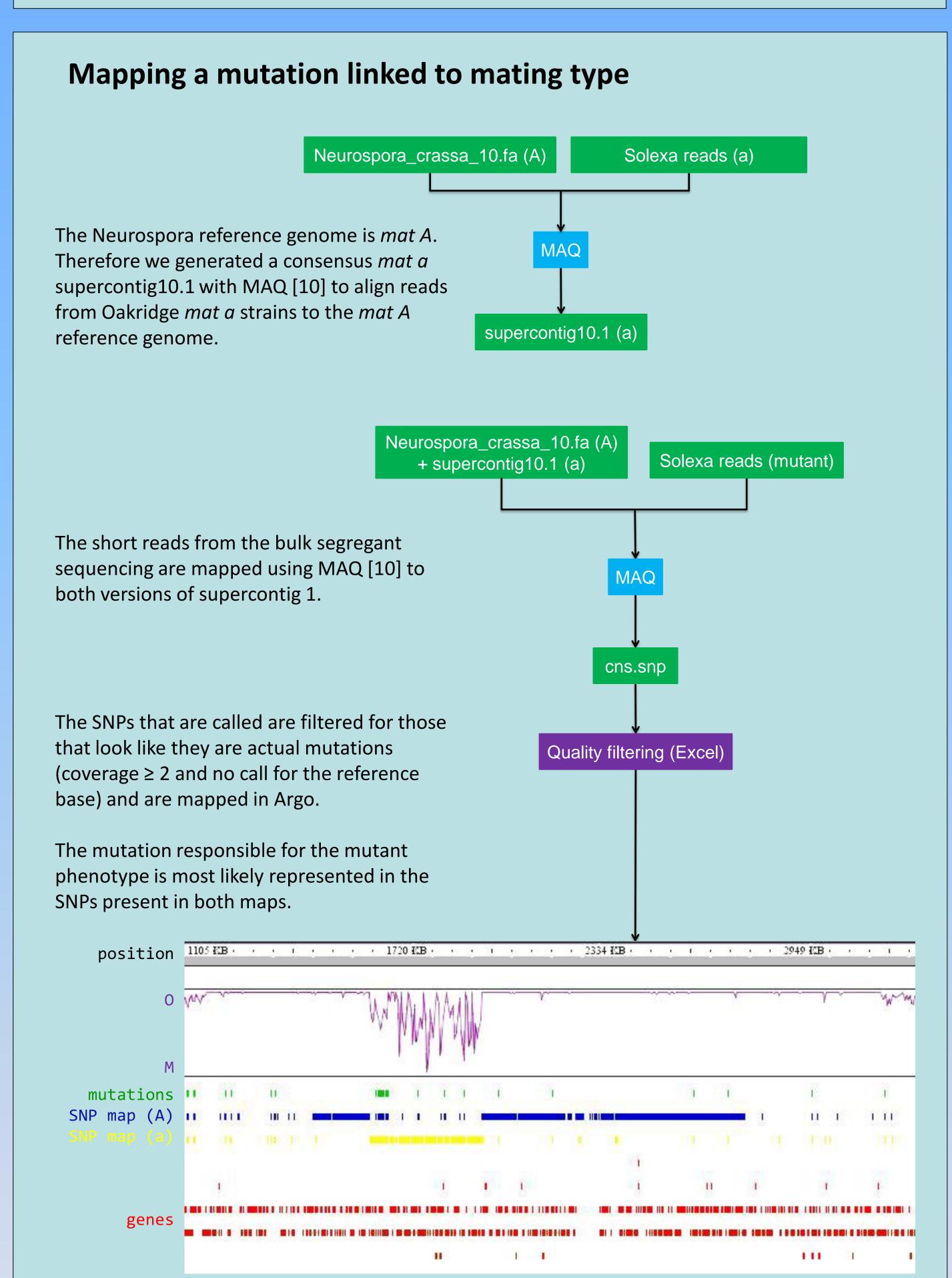


SNP profile in bulk segregants



Eighteen mat a progeny were obtained from a mapping cross between an Oakridge mutant (O) and Mauriceville (M). Their DNA was mixed in equal amounts, sequenced on a Solexa Genome Analyzer and mapped using the pipeline described.

Mapping a mutation by bulk segregant analysis Oakridge mutant Mauriceville Map mutation responsible for the curly phenotype by finding a peak of Oakridge SNPs when compared to a mixed OR/MV SNP background Oakridge Mauriceville Cross the strains. Pool the genomic DNA and sequence. (e) (e) Select mutant progeny and extract DNA.



Acknowledgements

We thank Mark Dasenko and Chris Sullivan for assistance with Illumina sequencing. We also thank the Neurospora Functional Genomics Project (NIH P01 GM068087) for primers and the Fungal Genetics Stock Center for strains. This work is funded by start-up funds from the Computational and Genome Biology Initiative at OSU and grants from the American Cancer Society (RSG-08-030-01-CCG) and DOE (DE-FG02-08ER64665).

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